

Review



Mentha arvensis and *Mentha* \times *piperita*-Vital Herbs with Myriads of Pharmaceutical Benefits

Hao Wei^{1,†}, Shuai Kong^{2,3,†}, Vanitha Jayaraman⁴, Dhivya Selvaraj⁵, Prabhakaran Soundararajan⁶ and Abinaya Manivannan^{7,*}

- ¹ School of Life Sciences, Qufu Normal University, Qufu 273165, China
- ² Power Technology Center, State Grid Shandong Electric Power Research Institute, Jinan 250002, China
- ³ College of Chemical and Biological Engineering, Zhejiang University, Hangzhou 310058, China
- ⁴ Faculty of Agriculture, SRM Institute of Science and Technology, Chengalpattu 603201, India
- ⁵ Department of Computer Science AI, Amrita School of Engineering, Amrita Vishwa Vidyapeeth, Chennai 601103, India
- ⁶ Evolutionary Developmental (Evo-Devo) Genetics Laboratory, National Institute of Plant Genome Research, Aruna Asaf Ali Marg, New Delhi 110067, India
- ⁷ National Institute of Plant Genome Research, Aruna Asaf Ali Marg, New Delhi 110067, India
 - Correspondence: abinayamanivannan@gmail.com
- + These authors contributed equally to this work.

Abstract: Mentha arvensis L. and Mentha × piperita L. are herbal plants belonging to the Lamiaceae family and are widely cultivated for their essential oils and culinary uses. These herbs are commercially valuable mints used in the preparation of herbal formulations, cosmetics, pharmaceuticals, and in food industries. Due to the presence of potential secondary metabolites, mints were employed to treat various disorders since ancient times in traditional medicines. The extracts of M. arvensis and M. \times *piperita* can improve the function of digestive system, central nervous system and respiratory system of the human body. Majority of the health benefits of these herbs are attributed by the essential oil components. In addition, the administration of M. arvensis and M. \times piperita under various pathological conditions studied in vitro and in vivo facilitated the recovery of detrimental ailments. Due to the increasing demand for natural product-based medicines, research is focused on the utilization of phytochemicals to treat various ailments. In order to provide a comprehensive overview of health benefits of M. arvensis and M. \times piperita, the present endeavor deals with the antioxidant property, anti-inflammatory property, anti-microbial, and anti-cancer activities of both species. However, a deeper knowledge on the specific metabolites of *M. arvensis* and *M. × piperita* and their mode of action against different disease targets will accelerate the discovery of novel natural drugs with less side effects and higher efficiency.

Keywords: Mentha; essential oil; pharmaceutical benefits; natural drugs

1. Introduction

Plants are viable reservoirs of phytochemicals with diverse medicinal values which form the vital ingredients for various pharmaceutical and herbal formulations [1]. Majority of the medicinal properties of plants are attributed by secondary metabolites. These metabolites render color, flavor, and aroma to plants which are utilized in food and drug industries [2]. Moreover, herbal plants are widely used in several cuisines as spices and condiments [3]. The chemo-diversity of secondary metabolites corresponds to different biological functions. According to World Health Organization (WHO), about 80% of population are interested on the utilization of traditional medicine which involves botanical isolates [4]. In recent times, research is ongoing particularly with respect to the gene or gene clusters associated with secondary metabolite biosynthesis [5] and organ specific accumulation of secondary metabolites with therapeutic values [6]. To facilitate the investigation of mint based bioactive compounds, the present endeavor provides a comprehensive



Citation: Wei, H.; Kong, S.; Jayaraman, V.; Selvaraj, D.; Soundararajan, P.; Manivannan, A. *Mentha arvensis* and *Mentha* × *piperita*-Vital Herbs with Myriads of Pharmaceutical Benefits. *Horticulturae* **2023**, *9*, 224. https://doi.org/10.3390/ horticulturae9020224

Academic Editors: Guillermo Cásedas, Cristina Moliner and Francisco Les

Received: 15 December 2022 Revised: 2 February 2023 Accepted: 2 February 2023 Published: 8 February 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). overview of secondary metabolites and nutraceutical effects of two commercially important mints such as *Mentha arvensis* L. and *Mentha* \times *piperita* L.

Mints are herbaceous plants with a collection of more than 60 species belonging to Lamiaceae family originated from Mediterranean basin and spread throughout the world by both natural and artificial means [7]. Mints are used as flavoring agents in food, beverage, chewing gums, and candies [8]. It is considered as healthy leafy vegetable because of its vitamins and mineral nutrients [9]. Due to the presence of potential secondary metabolites, mint was employed to treat various disorders since antiquity in ayurvedic and Chinese medicines [10]. The decoctions of mint aids in reduction of swelling, pain reliver, and is often used for the treatment of headache and eye redness, arthralgia, rubella, measles, chest, and hypochasm distension [10,11]. Numerous studies have evidenced that mint extracts can improve the digestive system, central nervous system, and respiratory system of the human body, and have anti-inflammatory antibacterial, antiviral, anticancer, antioxidation and other effects [12–14]. The abovementioned curing effects of mints are because of the occurrence of pharmaceutically valuable bioactive compounds. The main volatile components of mint include: menthol, menthone, menthyl acetate, menthofuran, and 1,8-cineol., etc. [15–18]. Non-volatile components present in mint are: flavonoids, phenolic acids, amino acids, nucleosides, terpenoids, etc., which are also the main active components of mint to play anti-inflammatory and antiviral effects [19,20]. The proportion and content of these components were affected by various factors such as growing environment, harvesting time, and variations in the species [21]. Among mint species, *M. arvensis* L. and M. × piperita L. are widely cultivated for the essential oil. M. arvensis is originated in the temperate climate of Europe, West Asia, and Central Asia [22]. $M. \times piperita$ is a natural hybrid between *M. viridis* and *M. aquatica* originated in the Mediterranean region [23]. The morphological differences of the two mint species are summarized in Table 1. Both mint species also displayed differences in content of essential oils. The essential oil of *M. arvensis* is made up of 30 to 50% menthol, 15 to 30% menthone, 3 to 10% menthyl acetate, and 1 to 5% other terpenes [24], whereas in $M \times piperita$, menthol accounted for about 36%, menthone 21%, menthyl acetate 7%, eucalyptol 7%, isomenthone 5%, neomenthol 4%, menthofuran 3%, D-limonene 2%, b-caryophyllene 2%, pulegone 1%, and b-pinene 1% [25]. Both mint species contributed to human health as excellent medicinal plants and raw materials for essential oil extraction.

Species	M. arvensis	M. $ imes$ piperita
Plant height	10–60 cm	30–100 cm
Stem	Purple at seedling stage and green at maturity, pilose	Purplish-red and glabrous
Leaf	Pale green, densely pilose along veins and sparsely puberulent elsewhere	Dark green, glabrous or below veins bristly and densely glandular
Flower	Corolla mauve, whorled on stems at leaf bases	Corolla white, lobes with pink halo, verticillaster form spica at the tips of stems and branches

Table 1. The morphological differences of *M. arvensis* and *M.* \times *piperita*.

The extracts of *M. arvensis* has been indulged in folklore medicine for the treatment of several diseases such as peptic ulcer, indigestion, skin allergies, respiratory diseases, etc. [26]. Various tissues of *M. arvensis* possess diverse medicinal properties. For instance, the decoctions obtained from leaf acts as diuretic agent, used to treat cold, asthma, rheumatism, jaundice, and other liver diseases [26]. *M. arvensis* oil, act as a natural source of anti-aflatoxigenic and antioxidant properties, protects the stored food commodities [27]. Furthermore, the essential oil provides a variety of health benefits, including antioxidant [28], anti-inflammatory [29], and antibacterial properties [30]. The essential oil from *M. arvensis* can serve as a natural pesticide [22]. Similarly, *M.* × *piperita* L. (peppermint) is a vital species in mint family with recognized pharmacological properties. It has been widely cultivated for its numerous benefits in agriculture, food, and medicinal purposes [31]. The essential oil of M. × *piperita* consists of myriads of bioactive phytochemicals with potential to treat several ailments [32–34]. The essential oils and secondary metabolites extracted from both mint species consisted of various health benefits.

2. Phytochemicals of *M. arvensis* and *M.* × *piperita*

Diverse bioactive phytochemicals were present in leaves, stem, and root tissues of *M. arvensis* [35]. The major components in *M. arvensis* are alkaloids, flavonoids, polyphenols, tannins, cardiac glycosides, and eugenol [35]. Flavonoids and terpenoids are mostly present in the leaves whereas diterpenes occur in both leaves and stems [35]. The M. arvensis consists of monoterpenes such as menthone, isomenthone, menthofuran, menthyl acetate cineole, limonene, and sesquiterpenes such as viridiflorol [22,24]. Flavonoids such as luteolin, menthoside, isorhoifolin, rutin hesperidin were also identified in *M. arvensis* [36]. Further, it also contains phenolic acids like caffeic acid, chlorogenic acid, and rosmarinic acid [37,38]. Triterpenes like squalene, α -amyrin, urosolic acid and sitosterol and phytol compounds like tocopherols, carotenoids, choline, betaine, cyclenes, rosmarinic acid, and tannin were also identified in *M. arvensis* [28]. Interestingly, a unique component linarin (acacetin-7-O- β -Drutionside) was detected in the flower tissue of *M. arvensis* [39]. Similarly, $M. \times piperita$ also consists of numerous phenolic compounds such as caffeic acid, rosmarinic acid, eriocitrin, luteolin-7-O-glucoside, gallic acid, p-Coumaric acid, chloregenic acid, sinapic acid, ellagic acid, hesperidin, and trans-ferulic acid [40–42]. Occurrence of flavones such as eriodictyol, naringenin, and hesperidin were identified in the leaf tissue of $M. \times piperita$ [43]. Leaf also consists of aglycones, palmitic acid, linoleic acid, linolenic acid, ascorbic acid, and soluble sugars [31]. Moreover, the presence of vital mineral nutrients such as potassium, calcium, sodium, phosphorus, zinc, and magnesium were also identified in the dried leaves of $M. \times piperita$ [44]. In general, the vital essential oil contents are present in the glandular trichomes of $M. \times piperita$ [11,45]. Phytochemical analysis of $M. \times piperita$ essential oil revealed the presence of several components such as menthone, iso-menthone, menthol, d-carvone, limonene, pulegone, and methyl petroselinate [46]. In addition, lignans and stilbenes were also identified in the extracts of M. \times *piperita* [11]. Taken together, the occurrence of bioactive phytochemicals with diverse beneficial properties makes both mint species an economically important choice for cultivation.

3. Uses of *M. arvensis* and *M.* \times *piperita*

Both Mentha species are widely utilized in food and beverage industries, cosmetics, agrochemicals, and herbal medicines due to presence of diverse phytochemicals [22,47,48]. The *Mentha* oil extracted from both species consists of a cooling effect, slight bitter taste, and strong aroma; due to these reasons, it can enhance the flavor and aroma of several products [48]. For instance, mint oil is used as the flavoring agent in candies, chewing gums, sauces, and beverages [47-50]. Moreover, the menthol-based cooling effect can act as a pain reliever which is a vital component in several pain-relieving balms and ointments [48–50]. Mentha-based dental products such as toothpastes and mouthwash causes a soothing cool effect in the oral cavities [49]. Further, Mentha oils are widely used essential oils for aromatherapy to relive stress and fatigue [51]. A recent study by Lin et al. [52] has demonstrated the changes occurring in electronencephalographic activity upon inhalation of peppermint oil under different visual stimuli. The results suggested that inhalation of peppermint oil enhanced the production of alpha waves in the pre-frontal area responsible for learning and thinking in white-sniffing group [52]. Additionally, the insect repellent property of mint oil is utilized to produce safer agro-chemicals and biopesticides [22]. The mint-based fodders are also employed in animal husbandry [53]. According to Masouri et al., [54] the supplementation of diet with peppermint oil increased the digestion of minerals and improved the meat quality in Japanese quails. Similarly, dietary supplementation of mint enhanced the egg laying capacity of hens [55]. Another wider usage of mint is in herbal medicines. Since antiquity mint is used to treat several ailments in various part of the world [46–50]. Mint are widely used to treat inflammation, bronchitis, toothaches, cramps, fever, headaches, and sore throat, etc. [46–49]. Moreover, the extracts of mint are employed in the treatment against intestinal colic, digestive disorders, ailments associated with gall bladders, gastric ulcers, and other gastrointestinal diseases [46–50]. The immense positive benefits of mints are widely attributed by its phytochemical constitutes with nutraceutical effects. However, some reports also illustrated the side effects of M. × *piperita* essential oil such as headache, vomiting, nausea, gastrointestinal discomforts, and heartburn [46–49]. This denotes a deeper knowledge on the mode of action of phytochemicals on disease targets is required. Therefore, in the future, research will focus on the development of novel mint based herbal formulation and identification of novel drug molecule with higher efficacy and lesser side effects has to be conducted.

4. Antioxidant Properties of *M. arvensis* and *M.* × *piperita*

Detoxification of harmful reactive oxygen species in higher concentrations generated in biological systems is inevitable for the proper physiological and metabolic functioning of an organism. The oxidative stress caused by these free radicals damages cellular proteins, lipids, and DNA molecules which results in diverse pathological conditions/diseases [56]. Natural compounds with antioxidant properties particularly from plant origin are always in demand. Plants consists of various phytochemicals with antioxidant compounds which facilitate the recovery from oxidative damages. These antioxidant property of plant extracts can be validated using various in vitro assays such as 2,2-diphenyl-1-picrylhydrazyl) (DPPH) radical scavenging, 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS⁺) analysis, and reducing power assays [47,57,58]. In most of these assays, the free radicals are propagated in a media which can be detoxified by the extract if it has antioxidant capacity, and this can be spectrophotometrically measured and compared with control like ascorbic acid. In general, polyphenolic compounds readily take part in the transfer of electrons and hydrogen atoms which attributes their antioxidant properties. To assess these potentials, the ability of the plant extract to reduce iron in the reaction mixture by the extracts are tested using various in vitro assays.

Apart from this mechanism, the phytochemicals can also act as chain terminators of free radicals making the unstable radicals into non-reactive stable compounds [43]. Several studies have evidenced the antioxidant properties of *M. arvensis* and $M. \times piperita$ using the invitro antioxidant assays. For instance, Dorman et al. [43] illustrated the antioxidant potentials of M. \times *piperita* extracted using different solvents which resulted in extraction efficiencies with different quantities and qualities. Among the solvents employed, methanolic extracts of M. \times *piperita* leaves displayed higher quantitative levels of total phenols and flavonoids. However, the significant qualitative antioxidants were observed in the ethanolic extracts of M. \times *piperita* with eriocitrin, rosmarinic acid, and naringenin-7-o-glucoside. Extracts of M. \times piperita displayed significant antioxidant property evidenced from increased reduction of iron(III), higher efficiency in DPPH, ABTS⁺, and H₂O₂ free radical scavenging properties [43]. Recent report by Hammad Al-Mijalli et al. [59] determined the volatile chemical compounds in M. \times *piperita* and evidenced its antioxidant properties. Likewise, the *M. arvensis* extracts illustrated significant free radical scavenging activities. Biswas et al. [28] validated antioxidant potentials of *M. arvensis* qualitatively using thin layer chromatography (TLC) and quantitatively by using DPPH assay. Moreover, the aqueous extracts of *M. arvensis* displayed varying levels of antioxidant properties in different assays such as DPPH, ABTS⁺, and phospholipid peroxidation assay [28]. Antioxidant properties of *M. arvensis* and *M.* \times *piperita* were exerted by various phytochemicals present in different tissues are listed in Table 2. Numerous research studies support the presence of free radical scavenging properties of *M. arvensis* and $M. \times piperita$, however, in order to proceed forward, studies unveiling the metabolite specific antioxidant mechanisms needs to be established. Further, molecular analysis of metabolic pathways responsible for the synthesis of potential antioxidants in both species will aid in the metabolic engineering of vital phytochemicals with antioxidant potentials. Moreover, a proper characterization

of antioxidant compounds and precise extraction approaches needs to be devised for the holistic utilization of the *Mentha* for large-scale production of antioxidant compounds.

Species	Tissue/Sample Extract	Class	Compound	References
M. arvensis	Essential oil	Terpenoid	Geranyl acetate Pulegone/Isopulegone Menthonel Isomenthone Menthyl acetate Menthyl acetate Menthol Menthol Menthyl acetate Pulegone Limonene Isomenthone Menthone Isomenthone	[60] [60] [60] [61] [61] [62] [61,63] [63] [63] [63] [63] [63] [63] [63]
			Menthol	[61,63]
M. × piperita	Rhizome	Phenolic acids	Protocatechuic aldehyde Luteolin-7-O-rutinoside Caffeic acid Salvianolic acid Lithospermic acid Salvianolic acid B Hesperetin-7-O-rutinoside Rosmarinic acid Eriodictyol-7-O-rutinoside	[64]
M. × piperita	Essential oil	Terpenoid	Sabinene β -Pinene β -Myrcene α -Terpinene Limonene 1,8-Cineole cis-Sabinene hydrate Linalool Menthone Menthofuran δ -Terpineol neo-Menthol Menthol Terpinen-4-ol Pulegone Piperitone Geranyl acetate (E)- β -Farnesene Germacrene D Elixene Viridiflorol Monoterpene Hydrocarbons Oxygenated monoterpenes Sesquiterpene hydrocarbons	[32]

Table 2. The phytochemicals with antioxidant potentials present in *M. arvensis* and *M.* \times *piperita*.

Species	Tissue/Sample Extract	Class	Compound	References
M. × piperita	Leaf	Phenols	Sinapic acid Gallic acid Catechin Caffeic acid Chloregenic acid Rutin Quercetin p-Coumaric acid Coumarin Carvacerol Vanilin Trans-ferulic acid Hesperedin Ellagic acid Eugenol Hesperetin	[65]
M. × piperita	Leaf	Flavanones	Naringenin Eriodictyiol Hesperetin Apigenin Luteolin Diosmetin	[60]

Table 2. Cont.

5. Anti-Inflammatory Activities of *M. arvensis* and *M.* \times *piperita*

In general, inflammation is a defense response to injury or infection to remove exogenous substances and facilitate self-healing. Inflammatory responses are triggered by a cascade of molecular mechanism and one of the main regulations is mediated by mitogen activated protein kinases (MAPKs)/nuclear factor-kappa B (NF-kB) signaling [66]. For this response, NF-kB is activated by mitogen activated protein kinases (MAPKs). Consequently, NF-kB can significantly influence the production of vital pro-inflammatory factors such as tumor necrosis factor- α (TNF- α), interleukin (IL-6), and IL-1 β [29]. Similarly, inflammatory reactions such as pain, fever, edema, and dysfunction can be mediated by nitric oxide (NO) and prostaglandin E_2 (PGE₂) signaling mediator regulated by inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 (COX-2) [67,68]. Essential oils (EOs) of M. arvensis and M. \times *piperita* have been evaluated for their anti-inflammatory activities in both in vivo or in vitro conditions. Recently, Kim et al. [29] reported the anti-inflammatory effects of *M. arvensis* essential oil in atopic dermatitis which is a chronic inflammatory skin disease. The molecular analysis revealed the essential oils of *M. arvensis* attenuated the inflammatory mediators in lipopolysaccharide (LPS)-stimulated RAW 264.7 macrophages. Further, the essential oils also inhibited phosphorylation of P65 and ERK necessary for activation of inflammatory response in NF-kB signaling pathway [29].

Similarly, in animal models, the immune-modulating effects of M. × *piperita* were evaluated. According to Atta et al. [68], pre-injection of the ethanol extract (200 and 400 mg/kg) of M. × *piperita* into mice significantly inhibited 49–50% acute inflammation of ear edema induced by topical xylene treatment. In another study, Mogosan et al. [69] identified that essential oils of M. × *piperita* attenuated the rat paw edema induced by λ -carrageenan. The in vitro anti-inflammatory effect was evaluated by measuring the secretion of proinflammatory cytokines including interleukin (IL)-1 β , TNF- α , IL-6, etc., or inflammatory mediators like NO and PGE₂ [69]. The EOs of M. × *piperita* and M. *arvensis* displayed similar mode of action to induce anti-inflammatory response by affecting the NF- κ B signaling pathway. For instance, Sun et al. [68] reported that M. × *piperita* essential oils effectively ceased NO and PGE₂ production in LPS-stimulated RAW 264.7 macrophages. A similar anti-inflammatory effect of essential oil of M. × *piperita* was also found in other LPS-stimulated cells, such as porcine alveolar macrophages (PAMs) [70] and HaCaT cells [29]. Menthol is the fundamental constitute of EOs derived from M. × *piperita* and M. *arvensis*, which significantly suppresses the production of TNF- α , IL-1 β , LTB4, and thromboxane B2 in LPS-stimulated human monocytes [71,72]. According to Juergens et al. [71], 1,8-cineole extracted from peppermint also possesses the ability to suppress the inflammatory mediating compounds [71]. Based on the above research, it can be conceived that essential oils of M. *arvensis* and M. × *piperita* exert repressing effects on cytokines or inflammatory mediators involved in the ERK/NF- κ B signaling pathway to provide an anti-inflammatory response (Figure 1). Thus, the essential oils of these mint species can be utilized for the herbal formulations associated with anti-inflammatory medicines.

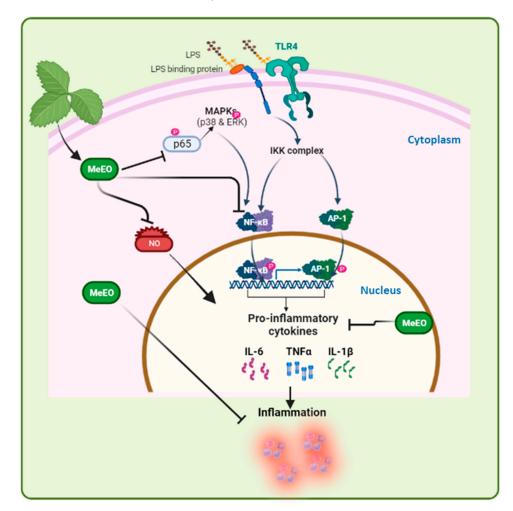


Figure 1. Schematic representation illustrating the anti-inflammatory activity exerted by *M. arvensis* and *M.* × *piperita* essential oils. MeEO, *Mentha* essential oil; LPS, Lipopolysaccharides; TLR4, Toll like receptor 4; MAPK, mitogen activated protein kinases; NF-kB, nuclear factor-kappa B; IKK, IkB kinase; AP1, Activator Protein 1, NO, nitric oxide; TNF- α , tumor necrosis factor- α ; The above figure is drawn using BioRender illustrations.

6. Anti-Bacterial Activity of *M. arvensis* and *M.* \times *piperita*

The antibacterial activities of M. × *piperita* and M. *arvensis* have been investigated extensively. The extracts can potentially inhibit the growth of various Gram-positive and Gram-negative bacteria (Table 3). Several reports illustrated the antibacterial activities of these extracts against pathogenic bacteria such as *Staphylococcus aureus* [30], *S. epidermidis* [73], *Salmonella typhimurium* [74], *Klebsiella pneumoniae* [75], etc. Moreover, studies have also compared the anti-bacterial activity of M. × *piperita* and M. *arvensis*, but the results were inconsistent. For instance, Hussain et al. [30] identified that the essential

oil of *M. arvensis* possess better anti-bacterial properties than M. \times *piperita*. For example, the *M. arvensis* displayed a minimum inhibitory concentration (MIC) of $20.3 \pm 1.0 \,\mu\text{g/mL}$ for *Bacillus subtilis*, whereas the MIC of $123.4 \pm 5.8 \,\mu\text{g/mL}$ was observed upon *M. piperita* treatment [30]. However, to the same strain, Heydari et al. [76] reported a MIC of 10 mg/mL with *M. arvensis* treatment while the MIC of M. \times *piperita* was reported as 0.1 mg/mL. This disparity in the results can be due to the differences in the chemical components of the extracts, plant materials from different geographical environment, seasonality, maturity of the plant, and the method of oil isolation [75–78]. In the studies above, higher percent of menthol were found in the essential oils with better antibacterial activities. Menthol, a cyclic monoterpene alcohol, has been confirmed with good antimicrobial activity [79–81]. It was identified as the major antimicrobial compound in M. \times piperita and M. arvensis oils [30,82]. The anti-bacterial mechanism of menthol has not been clearly elucidated, however it can be hypothesized as membrane-related inhibitory mechanisms [81]. Likewise, studies on the anti-bacterial function of terpenoids also suggested that their site of action was at the phospholipid bilayer, which would modify the membrane permeability and cause leakage of intracellular substances [83].

Further, Shalayel et al. [73] evaluated the anti-bacterial effect of $M_{\cdot} \times piperita$ extracted using different reagents, and discovered that the ethyl acetate extract had strongest anti-bacterial effects on the tested pathogens, followed by the chloroform, ethanol, and methanol extracts. On the other hand, Singh et al. [75] reported that ethyl acetate extract of $M. \times piperita$ possessed more inhibitory activity, indicating the primary importance of solvent used for extraction in determining the chemical composition. Additionally, Singh et al. [75] identified that the M. \times piperita oil is more effective against Gram-positive than Gram-negative bacteria. Similarly, Gochev et al. [84] evaluated the antimicrobial activity of Bulgarian peppermint oil and illustrated that peppermint oils are more effective against Gram-positive bacteria than Gram-negative bacteria. This might be due to the existence of lipopolysaccharides in the outer membrane of Gram-negative bacteria, which could enhance their resistance to antibacterial substances [82]. In addition, M. arvensis leaves displayed strong anti-bacterial effects against multidrug resistant Acinetobacter bauman*nii* [85]. The essential oil isolated from the fresh leaves of *M. arvensis* ceased the growth of five bacterial strains such as S. aureus, B. subtilis, S. pyogenes, E. coli, and P. aeruginosa, respectively [86,87]. The anti-bacterial activity of M. \times piperita and M. arvensis is due to the occurrence of mixture of phytochemicals particularly the essential oil components. In addition, based on the observed results, it can be assumed that the synergistic action of diverse phytochemicals in varying quantities present in the extracts could lead to the difference in the bactericidal activity of the extracts. However, the similar combinatorial activity of the phytochemical can also lead to lower effects against different bacterial strains. Therefore, future studies should be designed to delineate the primary constituent responsible for the anti-bacterial activity along with the molecular mechanism of action. Further, research on pathogen specific interaction anti-bacterial components in mints will facilitate its industrial and medicinal application.

Species	Sample Extract or Essential Oil	Origin of Bacterial Culture	Bacteria	^z MIC (mg/mL)	References
M. × piperita	Leaf extract	Pure cultures	Staphylococcus aureus ATCC 25923	12.0	
			Bacillus subtilis ATCC 10707	12.3	[30]
			Escherichia coli ATCC 25922	31.04	
M. arvensis	Leaf extract	Pure cultures	Staphylococcus aureus ATCC 25923	3.0.5	
			Bacillus subtilis ATCC 10707	2.03	[30]
			Escherichia coli ATCC 25922	33.03	

Table 3. Anti-bacterial activity of M. \times *piperita* and M. *arvensis*.

Species	Sample Extract or Essential Oil	Origin of Bacterial Culture	Bacteria	^z MIC (mg/mL)	References
			Staphylococcus aureus ATCC 9144	8.3	
M. × piperita	Essential oil		Enterococcus faecalis CIP 103907	8.3	[88]
			Listeria monocytogenes CRBIP 13.134	10	
		Pure cultures	Enterobacter aerogenes CIP 104725	>80	
			Escherichia coli CIP 105182	40	
			Pseudomonas aeruginosa CRBIP 19.249	>80	
			Salmonella enterica CIP 105150	8.3	
			Salmonella typhimurium ATCC 13311	13.3	
			Shigella dysenteria CIP 54.51	5.8	
			Escherichia coli <i>aDH5</i>	1.13	
		Pure cultures	Escherichia coli ATCC 25922	1.13	
			Pseudomonas aeruginosa	2.25	[00]
M. × piperita	Essential oil		Pseudomonas fluorescens	2.25	[89]
			Bacillus subtilis	1.13	
			Staphylococcus aureus	1.13	
			Staphylococcus aureus	62.67	
			Streptococcus pyogenes IBR S004	63.00	
			Streptococcus mutans IBR S001	60.33	
M. × piperita	Essential oil	Clinical oral	Lactobacillus acidophilus IBR L001	27.00	[90]
T T		isolates	Streptococcus salivarius IBR S006	34.66	[20]
			Streptococcus sanguinis IBR S002	28.66	
			Enterococcus faecalis IBR E001	23.86	
			Staphylococcus aureus	5.00	
			Staphylococcus epidermidis	2.50	
			Streptococcus pyogenes	1.25	[73]
	Extract of aerial part	a	Enterococcus faecalis	2.50	
		Clinical isolates	Escherichia coli	5.00	
M. imes piperita		from nosocomial	Klebsiella pneumoniae	10.00	
		patients	Pseudomonas aeruginosa	20.00	
			Seratia marcescens	10.00	
			Acinetobacter baumannii	40.00	
			Stenotrophomonas maltophilia	40.00	
	Essential oil	Pure cultures	Staphylococcus epidermidis	2.00	[76]
			Staphylococcus aureus	10.00	
M. imes piperita			Bacillus subtilis	0.10	
			Bacillus cereus	0.08	
	Essential oil	Pure cultures	Staphylococcus epidermidis	10.00	[76]
			Staphylococcus aureus	10.00	
M. arvensis			Bacillus subtilis	10.00	
			Bacillus cereus	10.00	
			Listeria monocytogenes PTCC 1163	12.50	
M. imes piperita	Essential oil	Pure cultures	Salmonella typhimurium ATCC 13311	25.00	[74]
		Clinical isolates	••		
M. arvensis	Essential oil	from vaginal	Streptococcus agalactiae ATCC 13813	1.80	[91]
11. 11 0011313	Looundar On	swabs	Lactobacillus spp.	1.80	[/+]

Table 3. Cont.

^z MIC = Minimum inhibitory concentration.

7. Antifungal Activity of *M. arvensis* and *M.* × *piperita*

The antifungal activities of *M. arvensis* and *M.* × *piperita* have been evidenced in various reports and most of the effects are due to the occurrence of essential oils. *Candida species* are opportunistic fungal pathogens to humans. Candidiasis caused by the infection of *Candida species* has a wide spectrum, varying from mild infection of skin or mucous membrane to severe invasion of organs [92]. Studies on the anti-fungal activities showed that the essential oils of *M.* × *piperita* and *M. arvensis* could inhibit the growth of *Candida species* and prevent the formation of biofilm [93–96]. The antifungal activity of *M.* × *piperita*

essential oil was comparable to that of amphotericin-B [93]. The formation of biofilm is the survival mode of *Candida species*, which can also contribute to its pathogenesis, and drug resistance. According to Saharkhiz et al. [95], the essential oil of M. \times *piperita* inhibited the biofilm formation of *Candida albicans* and *C. dubliniensis* at concentrations up to 2 µL/mL. The antifungal mechanism of M. \times *piperita* essential oil involves downregulation of the expression of some potential genes, such as secreted aspartyl proteinases (SAP 1, 2, 3, 9, 10) and hyphal wall protein 1 (HWP1) [93]. The essential oil of $M. \times piperita$ is also reported to be effective in inhibiting the growth of dermatophytes. The essential oils of M. \times *piperita* significantly ceased the growth of pathogenic fungi belonging to dermatophytes such as Trichophyton mentagrophytes, Microsporum canis, and M. gypseum [97–99]. The antimycotic effect of $M. \times piperita$ essential oil against Malassezia pachydermatis, a common cause of dermatitis in dogs were illustrated by Nardoni et al. [100]. Similarly, the antifungal activities of M. \times *piperita* against various plant pathogens have been widely studied. For instance, Rachitha et al. [101] discovered that $M. \times piperita$ essential oils showed inhibitory activity against Fusarium sporotrichioides, a filamentous fungi that contaminate corn and corn-based products. The application of M. \times *piperita* essential oil against the postharvest fungi Botrytis cinerea, Monilinia fructicola, Penicillium expansum, and Aspergillus niger identified to be effective against all tested fungi [102]. The inhibitory effect of $M. \times piperita$ essential oil against fungal pathogens of vegetables and mushrooms, including Botrytis cinerea, Sclerotinia sclerotiorum, Fusarium oxysporum, Phytophthora parasitica, Pythium aphanidermatum, Alternaria brassicae, Cladobotryum mycophilum, and Trichoderma aggressivum f.sp. europaeum were reported by Diánez et al. [103].

The antifungal property of *M. arvensis* essential oils enabled them to be applied in the field of food preservation. The augmentation of *M. arvensis* essential oil into gelatin edible coatings generated transparent film to prevent the infection of *Botrytis cinerea* and *Rhizopus stolonifera* in food and vegetable [104]. The ethanolic extracts of *M. arvensis* were assayed for antifungal activity against the strains of *C. albicans*, *C. tropicalis*, and *C. krusei*, and a potentiation effect was observed by Santos et al. [96]. The fungicidal activity for menthe, a volatile oil and principal constituent, has been established against Rhizoctonia solani and Fusarium moniliforme. It was also evaluated for antimycotic efficacy against F. oxysporum and *T. mentagrophytes* [105]. Till date, the mechanism of antifungal action by the essential oils are not revealed however earlier researches suggested that the mechanism can be due to the distortion of plasma membrane permeability of the fungus [106]. This result in the alteration of ion transport, respiration, and other vital metabolism of fungi. Overall, the above reports suggested efficient antifungal activity of *M. arvensis* and $M. \times piperita$ essential oils primarily mediated by alteration of plasma membrane permeability of the fungi as one of the modes of action. However, the occurrence of single or multiple metabolites with antifungal activity in *Mentha* require to be investigated. Further, existence of other possible mode of actions exerted by the bioactive phytochemicals in *Mentha* against pathogenic fungi needs to be elucidated.

8. Anticancer Activities of *M. arvensis* and *M.* \times *piperita*

Cancer is an increasingly serious health problem worldwide and has replaced heart disease as the leading cause of death worldwide [107]. Although the field of oncology medicine has made great progress in the present era, there are still several issues that need to be solved to improve cancer treatment. In the past decade, much research has focused on finding new therapies to reduce side effects caused by modern medicine [108]. Novel drug molecules with high efficiency, efficacy, less side effects and low environmental impact are desirable for the cancer treatment. To date, the search for the effective drug against most cancers are still in progress. Plant-based bioactive compounds provide opportunities for innovation in drug discovery. Several phytochemicals play an important role in the prevention and treatment of cancer. Among many phytochemicals with diverse structures, essential oils have attracted much attention due to their rich biological activities [109]. *M. arvensis* and *M.*× *piperita* also play important roles in the field of anticancer drug devel-

opment as major Mentha oil raw materials. Nano-emulsion of M. arvensis essential oil demonstrated high induction effect on early and middle apoptosis in anaplastic/aggressive thyroid cancer cells [110]. Likewise, methanol extract and essential oil from six different $M. \times piperita$ plants were effective against Vero, Hela, and Hep2 cancer cell lines [83]. In detail, the methanol extract has higher cytotoxic activities than essential oil for Vero and Hela whereas, the effect was reversed in Hep2 cancer cell line. These results indicated that there are differences in the effective components or mode of action in the extracts for cytotoxic activity on different types of cancer cells [111]. According to Jain et al. [112], the M. \times *piperita* extracts isolated with different organic solvents or water had different cytotoxic activity against six common human cancer cells. The results suggested that chloroform and ethyl acetate extracts induced significant dose-dependent and time-dependent anticancer activity, leading to G1 cell cycle arrest and mitochondria-mediated apoptosis, perturbation of oxidative balance, up-regulation of Bax genes, increased expression of p53 and P21, and induction of pro-inflammatory cytokine responses [112]. In most cases tumor formation is frequently accompanied by the development of new blood vessels. The extract of *M. arvensis* could facilitate the prevention of cancer by inhibiting the generation of blood vessels which was evidenced through chick chorioallantoic membrane assay [113]. In another study, $M \times piperita$ extract not only restrained skin papilloma by influencing the activation/detoxification of the carcinogen but also improved cellular resistance to radiation through enhancing the antioxidant mechanism [114]. Likewise, M. arvensis extract suppressed the growth and induced apoptosis against HepG2 cell lines which was observed by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) assay [47]. Moreover, the *M. arvensis* also displayed anti-cancer activity against eight human cancer cells from six different origins such as breast, colon, prostate, lung, leukemia, and glioblastoma [47]. Further, green silver nanoparticles synthesized from *M. arvensis* displayed a promising role against the breast cancer cell line with less toxic to normal cells, non-mutagenic, mediated caspase 9-dependent apoptosis in MCF7 and MDA-MB-231 cells [115]. An essential oil which contains menthol, menthone has anticancer property in treating anaplastic thyroid cancer cells by nanoemulsion method [110]. According to Weecharangsan et al. [116], essential oils of *M. arvensis* showed cytotoxic activity with IC_{50} value of 142.0 \pm 92.2 and 243.3 \pm 151.1 μ g/mL against KB cells and with IC₅₀ value of 178.4 \pm 92.1 and 779.4 \pm 673.3 μ g/mL against HeLa cells. Moreover, the *M. arvensis* extract contains Rosmarinic acid(Ra), a natural phenolic compound which induced dosedependent cell death in cultured HepG2 cells and mRNA expression analysis confirmed the down regulation of anti-apoptotic gene Bcl2 with upregulation of proapoptotic genes like Bax and ERK2 [117]. Similar to other aromatic herb extracts, monoterpenes, sesquiterpenes, oxygenated monoterpenes, oxygenated sesquiterpenes, and phenolics, among others, play key roles in the anticancer mechanism of mint extract. The main mechanism of chemoprophylaxis of these compounds is protection against oxidation, mutation and proliferation, enhancement of immune function and monitoring, enzyme induction and improvement of detoxification ability, regulation of multidrug resistance, and synergistic action. However, more research on the identification of specific components and their mechanism of action needs to be performed to promote the treatment of cancer by natural products such as mint extracts in the future.

9. Extracts of *M. arvensis* and *M.* × *piperita* in Radiation Therapy

Radiotherapy is a vital method for the treatment of malignant tumors. About 50% of cancer patients need radiation therapy in the course of cancer treatment, and about 40% of cancers can be suppressed with radiation [118]. At present, targeted radiations towards cancerous cells without affecting the normal cells is an immensely researched area to provide effective anti-cancer treatment [119]. In the chemical radiation therapy, sensitizers or protectors possess the ability to guard the normal cells to an extent from the damages caused by radiations [120]. However, the radiation protectors discovered earlier, such as 2-(3-aminopropylamino) ethylsulphanyl phosphonic acid (WR-2721), are toxic to

cells at the optimal protective concentration [121]. Therefore, research has been focused on the identification of effective plant/natural-product-based radiation protectors for cancer treatments. One of the promising alternates is the extract of *M. arvensis*. According to Jagetia and Baliga [122], mice fed with different doses of *M. arvensis* extract had significantly decreased the side effects of gamma radiations in comparison with the control. The results showed that the optimal protection was provided by feeding 10 mg/kg *M. arvensis* extract, which was much lower than the toxic dose (1000 mg/kg). Similarly, radiation protection function was also identified in M. × *piperita* [123–125]. Under in vivo conditions in the mice model, M. \times *piperita* extract significantly improved serum alkaline phosphatase and declined serum acid phosphatase activities. Alkaline and acid phosphatase plays an important role in the maintenance of cell permeability [126–128]. These reports demonstrated that $M. \times piperita$ alleviates the damages to animal cell membranes caused by radiation to some extent. Likewise, $M. \times piperita$ extract aided in the recovery of intestinal mucosa [124], bone marrow [125], and testicles [129] from radiation damages in mice. This illustrates that Mentha extract consists of promising phytochemicals which can prevent the cellular damages caused by radiations. It is apparent from the above studies that the extracts of M. *arvensis* and M. \times *piperita* can act as vital components for recovery from damages caused by radiations but the exact rationale behind the anti-radiation property needs to be identified.

10. Conclusions and Future Perspectives

Versatile nutraceutical potentials of *M. arvensis* and $M. \times piperita$ have been evidenced from various studies denoting their enormous pharmaceutical importance, most of which has been attributed to its antioxidant, anti-microbial, anti-cancerous, and anti-inflammatory properties. The administration of *M. arvensis* and $M. \times piperita$ under various pathological conditions facilitate the recovery of diseases and aids in the prevention of detrimental ailments. Particularly, the occurrence of volatile essential oils in leaves, stems, and roots consists of the disease combating potentials which can be utilized for discovery of novel drugs with higher efficacy and lesser side effects. Even though mint consists of numerous economic and health benefits, to date, there is a scarcity of knowledge in the aspects of quality and quantity of the phytochemicals and choice of genotypes to be used as raw materials for large-scale industrial usage. In addition, the presence of excessive flavor or aroma in mint can negatively influence the organoleptic traits and reduce the consumer quality of the products. Therefore, research needs to address the abovementioned shortcomings to enhance the elite varieties of mint with optimal phytochemicals. Further, clinical trials with Mentha for diverse disorders should be initiated to validate the potential of the natural herbal medicines in human. Additionally, appropriate channel of knowledge on the method of consumption, elucidation of molecular mechanism behind the antioxidant, anti-cancer, and anti-microbicidal effects of M. arvensis and M. \times piperita needs to be devised. In addition, application of current omics based approaches to unveil the metabolic pathways and candidate genes involved in the essential oil biosynthesis will facilitate the utilization of these *Mentha* species in large-scale in food and pharmaceutical fields.

Author Contributions: Conceptualization, A.M. and H.W.; investigation, S.K.; writing—original draft preparation, H.W., S.K., V.J. and D.S.; writing—review and editing, P.S. and A.M.; supervision, A.M.; funding acquisition, A.M. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Informed Consent Statement: Not applicable.

Acknowledgments: Abinaya Manivannan, DST-INSPIRE faculty (DST/INSPIRE/04/2021/003731) acknowledges the support from Department of Science and Technology (DST), Government of India.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Süntar, I. Importance of ethnopharmacological studies in drug discovery: Role of medicinal plants. *Phytochem. Rev.* **2020**, *19*, 1199–1209.
- 2. Hassan, B. Medicinal plants (importance and uses). *Pharm. Anal. Acta.* **2012**, *3*, e139.
- Li, Y.; Kong, D.; Fu, Y.; Sussman, M.R.; Wu, H. The effect of developmental and environmental factors on secondary metabolites in medicinal plants. *Plant Physiol. Biochem.* 2020, 148, 80–89. [CrossRef]
- 4. Hosseinzadeh, S.; Jafarikukhdan, A.; Hosseini, A.; Armand, R. The Application of Medicinal Plants in Traditional and Modern Medicine: A Review of Thymus vulgaris. *Int. J. Clin. Med.* **2015**, *6*, 635–642. [CrossRef]
- Alami, M.M.; Ouyang, Z.; Zhang, Y.; Shu, S.; Yang, G.; Mei, Z.; Wang, X. The Current Developments in Medicinal Plant Genomics Enabled the Diversification of Secondary Metabolites' Biosynthesis. *Int. J. Mol. Sci.* 2022, 23, 15932. [CrossRef] [PubMed]
- 6. Mondal, M.; Chandra, I. Organ specific phytochemical changes and antioxidant activities of in vivo and in vitro grown *Gloriosa superba* L. *S. Afr. J. Bot.* **2023**, 152, 1–10.
- Tucker, A.O. Mentha: Economic uses. In *Mint: The Genus Mentha*, 1st ed.; Lawrence, B.M., Ed.; CRC Press: Boca Raton, FL, USA, 2006; pp. 519–528.
- Ćavar Zeljković, S.; Šišková, J.; Komzáková, K.; De Diego, N.; Kaffková, K.; Tarkowski, P. Phenolic Compounds and Biological Activity of Selected Mentha Species. *Plants* 2021, 10, 550.
- Arzani, A.; Zeinali, H.; Razmjo, K. Iron and magnesium concentrations of mint accessions (*Mentha* spp.). *Plant Physiol. Biochem.* 2007, 45, 323–329.
- 10. Liu, Y.; Wang, Z.; Zhang, J. Dietary Chinese Herbs, 1st ed.; Springer: Vienna, Austria, 2015; pp. 631–636.
- 11. Mahendran, G.; Rahman, L. Ethnomedicinal, phytochemical and pharmacological updates on Peppermint (*Mentha* × *piperita* L.)— A review. *Phytother. Res.* **2020**, *34*, 2088–2213. [CrossRef] [PubMed]
- 12. Balakrishnan, A. Therapeutic uses of peppermint—A review. J. Pharm. Sci. Res. 2015, 7, 474–476.
- 13. McKay, D.L.; Blumberg, J.B. A review of the bioactivity and potential health benefits of peppermint tea (*Mentha piperita* L.). *Phytother. Res.* **2006**, *20*, 619–633.
- 14. Spirling, L.I.; Daniels, I.R. Botanical perspectives on health peppermint: More than just an after-dinner mint. *J. R. Soc. Promot. Health* **2001**, *121*, *62–63*. [PubMed]
- 15. Beigi, M.; Torki-Harchegani, M.; Ghasemi Pirbalouti, A. Quantity and chemical composition of essential oil of peppermint (*Mentha* × *piperita* L.) leaves under different drying methods. *Int. J. Food Prop.* **2018**, *12*, 267–276.
- 16. Mahboubi, M.; Kazempour, N. Chemical composition and antimicrobial activity of peppermint (*Mentha piperita* L.) Essential oil. *Songklanakarin J. Sci. Technol.* **2014**, *36*, 83–87.
- 17. Sivropoulou, A. Antimicrobial activity of mint essential oils. J. Agric. Food Chem. 1995, 43, 2384–2388. [CrossRef]
- Tsai, M.; Wu, C.T.; Lin, T.F.; Lin, W.; Huang, Y.C.; Yang, C.H. Chemical Composition and Biological Properties of Essential Oils of Two Mint Species. Trop. J. Pharm. Res. 2013, 12, 577–582. [CrossRef]
- 19. Jayanthy, G.; Subramanian, S. Extraction, Isolation and Characterization of Rosmarinic Acid, a Major Polyphenol in Non-volatile Constituent of Mint Leaves. *Asian J. Res. Chem.* **2013**, *6*, 1160–1165.
- Xu, L.-L.; Xu, J.-J.; Zhong, K.-R.; Shang, Z.-P.; Wang, F.; Wang, R.-F.; Zhang, L.; Zhang, J.-Y.; Liu, B. Analysis of Non-Volatile Chemical Constituents of Menthae Haplocalycis Herba by Ultra-High Performance Liquid Chromatography-High Resolution Mass Spectrometry. *Molecules* 2017, 22, 1756. [PubMed]
- 21. Grulova, D.; De Martino, L.; Mancini, E.; Salamon, I.; De Feo, V. Seasonal variability of the main components in essential oil of *Mentha* × *piperita* L. *J. Sci. Food Agric.* **2015**, *95*, 621–627.
- 22. Kalemba, D.; Synowiec, A. Agrobiological interactions of essential oils of two menthol mints: Mentha piperita and Mentha arvensis. *Molecules* **2019**, *25*, 59. [CrossRef]
- 23. Bacon, F.J. The botanical origin of American peppermint-Mentha piperita L. J. Am. Pharm. Assoc. 1928, 17, 1094-1096. [CrossRef]
- 24. Singh, A.; Raina, V.; Naqvi, A.; Patra, N.; Kumar, B.; Ram, P.; Khanuja, S. Essential oil composition and chemoarrays of menthol mint (*Mentha arvensis* L. f. *piperascens* Malinvaud ex. Holmes) cultivars. *Flavour Fragr. J.* 2005, 20, 302–305. [CrossRef]
- Skalicka-Woźniak, K.; Walasek, M. Preparative separation of menthol and pulegone from peppermint oil (*Mentha piperita* L.) by high-performance counter-current chromatography. *Phytochem. Lett.* 2014, 10, 94–98.
- Akram, M.; Uzair, M.; Malik, N.S.; Mahmood, A.; Asif, H.M. Mentha arvensis Linn.: A review article. J. Med. Plant Res. 2011, 5, 4499–4503.
- 27. Kumar, A.; Shukla, R.; Singh, P.; Singh, A.K.; Dubey, N.K. Use of essential oil from *Mentha arvensis* L. to control storage moulds and insects in stored chickpea. J. Sci. Food Agric. 2010, 89, 2643–2649. [CrossRef]
- Biswas, N.N.; Saha, S.; Ali, M.K. Antioxidant, antimicrobial, cytotoxic and analgesic activities of ethanolic extract of *Mentha* arvensis L. Asian Pac. J. Trop. Biomed. 2014, 4, 792–797.
- Kim, S.Y.; Han, S.D.; Kim, M.; Mony, T.J.; Lee, E.S.; Kim, K.M.; Choi, S.H.; Hong, S.H.; Choi, J.W.; Park, S.J. Mentha arvensis Essential Oil Exerts Anti-Inflammatory in LPS-Stimulated Inflammatory Responses via Inhibition of ERK/NF-κB Signaling Pathway and Anti-Atopic Dermatitis-like Effects in 2,4-Dinitrochlorobezene-Induced BALB/c Mice. *Antioxidants* 2021, 10, 1941. [CrossRef]
- Hussain, A.I.; Anwar, F.; Nigam, P.S.; Ashraf, M.; Gilani, A.H. Seasonal variation in content, chemical composition and antimicrobial and cytotoxic activities of essential oils from four Mentha species. J. Sci. Food Agric. 2010, 90, 1827–1836. [CrossRef]

- 31. Gholamipourfard, K.; Salehi, M.; Banchio, E. Review: Mentha piperita phytochemicals in agriculture, food industry and medicine: Features and applications. *S. Afr. J. Bot.* **2021**, *141*, 183–195. [CrossRef]
- Ahmadi, H.; Morshedloo, M.R.; Emrahi, R.; Javanmard, A.; Rasouli, F.; Maggi, F.; Kumar, M.; Lorenzo, J.M. Introducing Three New Fruit-Scented Mints to Farmlands: Insights on Drug Yield, Essential-Oil Quality, and Antioxidant Properties. *Antioxidants* 2022, 11, 866. [CrossRef] [PubMed]
- Zhao, H.; Ren, S.; Yang, H.; Tang, S.; Guo, C.; Liu, M.; Tao, Q.; Ming, T.; Xu, H. Peppermint essential oil: Its phytochemistry, biological activity, pharmacological effect and application. *Biomed. Pharmacother.* 2022, 154, 113559.
- 34. Sahandi, M.S.; Mehrafarin, A.; Khalighi-Sigaroodi, F.; Sharifi, M.; Badi, H.N. Review on anatomical, phytochemical and pharmacological properties of peppermint (*Mentha piperita* L.). J. Med. Plants **2018**, 17, 16–33.
- Farnaz, M.; Shahzad, H.; Alia, S.; Ghazala, P.; Amina, W.; Shazia, S.; Rafique, A.C.; Rashid, M.; Humayun, R.; Muhammad, I.; et al. Phyto-chemical analysis, anti-allergic and anti-inflammatory activity of Mentha arvensis in animals. *Afr. J. Pharm. Pharmacol.* 2012, *6*, 613–619.
- 36. Thawkar, B.S. Phytochemical and pharmacological review of Mentha arvensis. Int. J. Green Pharm. 2016, 10, 71–76.
- Mahishi, P.; Srinivasa, B.H.; Shivanna, M.B. Medicinal plant wealth of local communities in some villages in Shimoga District of Karnataka, India. J. Ethnopharmacol. 2005, 98, 307–312.
- Kumar, A.; Chattopadhyay, S. DNA damage protecting activity and antioxidant potential of pudina extract. *Food Chem.* 2007, 100, 1377–1384. [CrossRef]
- Oinonen, P.P.; Jokela, J.K.; Hatakka, A.I.; Vuorela, P.M. Linarin, a selective acetylcholinesterase inhibitor from Mentha arvensis. *Fitoterapia* 2006, 77, 429–434.
- Trevisan, S.C.C.; Menezes, A.P.P.; Barbalho, S.M.; Guiguer, É.L. Properties of mentha piperita: A brief review. World J. Pharm. Med. Res. 2017, 3, 309–313.
- Badal, R.M.; Badal, D.; Badal, P.; Khare, A.; Shrivastava, J.; Kumare, V. Pharmacological Action of Mentha piperita on Lipid Profile in Fructose-Fed Rats. *Iran. J. Pharm. Res.* 2011, 10, 843–848.
- 42. Patil, S.R.; Patial, R.S.; Godghate, A.G. *Mentha piperita* Linn: Phytochemical, antibacterial and dipterian adulticidal approach. *Int. J. Pharm. Pharm. Sci.* **2016**, *8*, 352–355.
- Dorman, H.J.; Koşar, M.; Başer, K.H.; Hiltunen, R. Phenolic profile and antioxidant evaluation of *Mentha* × *piperita* L. (peppermint) extracts. *Nat. Prod. Commun.* 2009, 4, 535–542. [PubMed]
- 44. Uribe, E.; Marín, D.; Vega-Gálvez, A.; Quispe-Fuentes, I.; Rodríguez, A. Assessment of vacuum-dried peppermint (*Mentha piperita* L.) as a source of natural antioxidants. *Food Chem.* **2016**, *190*, 559–565. [PubMed]
- 45. Qamar, N.; Pandey, M.; Vasudevan, M.; Kumar, A.; Shasany, A.K. Glandular trichome specificity of menthol biosynthesis pathway gene promoters from *Mentha* × *piperita*. *Planta*. **2022**, *256*, 110. [CrossRef] [PubMed]
- Brahmi, F.; Khodir, M.; Mohamed, C.; Pierre, D. Chemical composition and biological activities of Mentha species. In *Aromatic and Medicinal Plants-Back to Nature*, 1st ed.; EI-Shemy, H.A., Ed.; InTech: Rijeka, Croatia, 2017; Volume 10, pp. 47–79.
- Tafrihi, M.; Imran, M.; Tufail, T.; Gondal, T.A.; Caruso, G.; Sharma, S.; Sharma, R.; Atanassova, M.; Atanassov, L.; Valere Tsouh Fokou, P.; et al. The wonderful activities of the genus Mentha: Not only antioxidant properties. *Molecules* 2021, 26, 1118. [CrossRef] [PubMed]
- Saqib, S.; Ullah, F.; Naeem, M.; Younas, M.; Ayaz, A.; Ali, S.; Zaman, W. Mentha: Nutritional and Health Attributes to Treat Various Ailments Including Cardiovascular Diseases. *Molecules* 2022, 27, 6728. [CrossRef]
- 49. Anwar, F.; Abbas, A.; Mehmood, T.; Gilani, A.H.; Rehman, N.U. Mentha: A genus rich in vital nutra-pharmaceuticals—A review. *Phytother. Res.* **2019**, *33*, 2548–2570.
- 50. Eftekhari, A.; Khusro, A.; Ahmadian, E.; Dizaj, S.M.; Hasanzadeh, A.; Cucchiarini, M. Phytochemical and nutra-pharmaceutical attributes of Mentha spp.: A comprehensive review. *Arab. J. Chem.* **2021**, *14*, 103106.
- 51. Kiełtyka-Dadasiewicz, A.; Kubat-Sikorska, A. Chemical diversity of mint essential oils and their significance for aromatherapy. *Arch. Physiother. Glob. Res.* **2018**, *22*, 53–59. [CrossRef]
- 52. Lin, S.; Wang, Y.; Wu, K.; Yu, G.; Liu, C.; Su, C.; Yi, F. Study on the Effect of *Mentha* × *piperita* L. Essential Oil on Electroencephalography upon Stimulation with Different Visual Effects. *Molecules* **2022**, *27*, 4059.
- Horky, P.; Skalickova, S.; Smerkova, K.; Skladanka, J. Essential oils as a feed additives: Pharmacokinetics and potential toxicity in monogastric animals. *Animals* 2019, 9, 352. [CrossRef]
- 54. Masouri, L.; Bagherzadeh-Kasmani, F.; Mehri, M.; Rokouei, M.; Masouri, B. Mentha piperita as a promising feed additive used to protect liver, bone, and meat of Japanese quail against aflatoxin B1. *Trop. Anim. Health Prod.* **2022**, *54*, 254.
- 55. Abdel-Wareth, A.A.A.; Lohakare, J.D. Effect of dietary supplementation of peppermint on performance, egg quality, and serum metabolic profile of Hy-Line Brown hens during the late laying period. *Anim. Feed Sci. Technol.* **2014**, *197*, 114–120. [CrossRef]
- Farnad, N.; Heidari, R.; Aslanipour, B. Phenolic composition and comparison of antioxidant activity of alcoholic extracts of Peppermint (*Mentha piperita*). J. Food Meas. Charact. 2014, 8, 113–121. [CrossRef]
- 57. Anwar, F.; Alkharfy, K.M.; Najeeb-ur-Rehman; Adam, E.; Gilani, A. Chemo-geographical Variations in the Composition of Volatiles and the Biological Attributes of *Mentha longifolia* (L.) Essential Oils from Saudi Arabia. *Int. J. Pharmacol.* 2017, 13, 408–424.
- Nickavar, B.; Alinaghi, A.; Kamalinejad, M. Evaluation of the Antioxidant Properties of Five Mentha Species. Iran. J. Pharm. Res. 2008, 7, 203–209.

- 59. Hamad Al-Mijalli, S.; Elsharkawy, E.; Abdallah, E.; Hamed, M.; El Omari, N.; Mahmud, S.; Alshahrani, M.; Naceiri Mrabti, H.; Bouyahya, A. Determination of Volatile Compounds of *Mentha piperita* and *Lavandula multifida* and Investigation of Their Antibacterial, Antioxidant, and Antidiabetic Properties. *Evid. Based Complement. Alternat. Med.* 2022, 2022, 9306251.
- 60. Mimica-Dukic, N.; Bozin, B. *Mentha* L. species (Lamiaceae) as promising sources of bioactive secondary metabolites. *Curr. Pharm. Des.* **2008**, *14*, 3141–3150. [CrossRef]
- Kumar, A.; Khajuria, V.; Aggarwal, S. Secondary metabolites of *Mentha arvensis* and their biological activities. *Anal. Chem. Lett.* 2012, 2, 373–400. [CrossRef]
- 62. Phatak, S.V.; Heble, M.R. Organogenesis and terpenoid synthesis in Mentha arvensis. Fitoterapia 2002, 73, 32–39. [CrossRef]
- 63. Venkatesha, K.T.; Padalia, R.; Singh, V.R.; Upadhyay, R.K.; Chauhan, A. Correlation and path-analysis for morpho-economic traits and chemical constituents of essential oil in Corn mint (*Mentha arvensis* L.) accessions. *Arab. J. Med. Aromat. Plants* **2020**, *6*, 1–16.
- 64. Bittner Fialová, S.; Kurin, E.; Trajčíková, E.; Jánošová, L.; Šušaníková, I.; Tekeľová, D.; Nagy, M.; Mučaji, P. Mentha Rhizomes as an Alternative Source of Natural Antioxidants. *Molecules* **2020**, *25*, 200. [PubMed]
- Mahdavikia, F.; Saharkhiz, M.J. Phytotoxic activity of essential oil and water extract of peppermint (*Mentha* × *piperita* L. CV. Mitcham). J. Appl. Res. Med. Aromat. Plants 2015, 2, 146–153. [CrossRef]
- 66. Liu, T.; Zhang, L.; Joo, D.; Sun, S.C. NF-κB signaling in inflammation. Signal Transduct. Target. Ther. 2017, 2, 17023. [CrossRef]
- Won, J.H.; Kim, J.Y.; Yun, K.J.; Lee, J.H.; Back, N.I.; Chung, H.G.; Chung, S.A.; Jeong, T.S.; Choi, M.S.; Lee, K.T. Gigantol isolated from the whole plants of Cymbidium goeringii inhibits the LPS-induced iNOS and COX-2 expression via NF-kappaB inactivation in RAW 264.7 macrophages cells. *Planta Med.* 2006, 72, 1181–1187. [CrossRef] [PubMed]
- Zamora, R.; Vodovotz, Y.; Billiar, T.R. Inducible Nitric Oxide Synthase and Inflammatory Diseases. *Mol. Med.* 2000, *6*, 347–373. [PubMed]
- 69. Atta, A.H.; Alkofahi, A. Anti-nociceptive and anti-inflammatory effects of some Jordanian medicinal plant extracts. *J. Ethnopharmacol.* **1998**, *2*, 117–124. [CrossRef]
- Hejna, M.; Kovanda, L.; Rossi, L.; Liu, Y. Mint Oils: In Vitro Ability to Perform Anti-Inflammatory, Antioxidant, and Antimicrobial Activities and to Enhance Intestinal Barrier Integrity. *Antioxidants* 2021, 10, 1004.
- Juergens, U.R.; Stöber, M.; Vetter, H. The anti-inflammatory activity of L-menthol compared to mint oil in human monocytes in vitro: A novel perspective for its therapeutic use in inflammatory diseases. *Eur. J. Med. Res.* 1998, 3, 539–545.
- 72. Hilfiger, L.; Triaux, Z.; Marcic, C.; Héberlé, E.; Emhemmed, F.; Darbon, P.; Marchioni, E.; Petitjean, H.; Charlet, A. Anti-Hyperalgesic Properties of Menthol and Pulegone. *Front. Pharmacol.* **2021**, *12*, 3248. [CrossRef]
- 73. Shalayel, M.H.F.; Asaad, A.M.; Qureshi, M.A.; Elhussein, A.B. Anti-bacterial activity of peppermint (Mentha piperita) extracts against some emerging multi-drug resistant human bacterial pathogens. *J. Herb. Med.* **2017**, *7*, 27–30.
- Raeisi, M.; Hashemi, M.; Ansarian, E.; Hejazi, J.; Hassanzadazar, H.; Daneshamooz, S.; Jannat, B.; Aminzare, M. Antibacterial Effect of Mentha piperita Essential Oil Against Foodborne Pathogens in Minced Meat During Storage at Abuse Refrigeration Temperature. *Adv. Anim. Vet. Sci.* 2019, 7, 720–726.
- 75. Singh, R.; Shushni, M.; Belkheir, A. Antibacterial and antioxidant activities of Mentha piperita L. Arab. J. Chem. 2015, 8, 322–328.
- 76. Heydari, M.; Zanfardino, A.; Taleei, A.; Bushehri, A.A.S.; Hadian, J.; Maresca, V.; Sorbo, S.; Napoli, M.D.; Varcamonti, M.; Basile, A.; et al. Effect of Heat Stress on Yield, Monoterpene Content and Antibacterial Activity of Essential Oils of *Mentha* × *piperita* var. Mitcham and *Mentha arvensis* var. piperascens. *Molecules* 2018, 23, 1903. [CrossRef]
- Bui-Phuc, T.; Nhu-Trang, T.T.; Cong-Hau, N. Comparison of chemical composition of essential oils obtained by hydro-distillation and microwave-assisted extraction of Japanese mint (*Mentha arvensis* L.) grown in Vietnam. *IOP Conf. Ser. Mater. Sci. Eng.* 2020, 991, 012039. [CrossRef]
- 78. Ibrahim, M.; Ankwai, G.; Gungshik, J.; Taave, P. Comparative extraction of essential oils of *Mentha piperita* (mint) by steam distillation and enfleurage. *Niger. J. Chem. Res.* 2022, 26, 2021. [CrossRef]
- 79. Osawa, K.; Saeki, T.; Yasuda, H.; Hamashima, H.; Sasatsu, M.; Arai, T. The antibacterial activities of peppermint oil and green tea polyphenols, alone and in combination, against Enterohemorrhagic Escherichia coil. *Biocontrol Sci.* **1999**, *4*, 1–7.
- Pattnaik, S.; Subramanyam, V.R.; Bapaji, M.; Kole, C.R. Antibacterial and antifungal activity of aromatic constituents of essential oils. *Microbios* 1997, 89, 39–46. [PubMed]
- 81. Trombetta, D.; Castelli, F.; Sarpietro, M.G.; Venuti, V.; Cristani, M.; Daniele, C.; Saija, A.; Mazzanti, G.; Bisignano, G. Mechanisms of antibacterial action of three monoterpenes. *Antimicrob. Agents Chemother.* **2005**, *49*, 2474–2478. [CrossRef]
- 82. Işcan, G.; Kirimer, N.; Kürkcüoğlu, M.; Başer, K.H.; Demirci, F. Antimicrobial screening of *Mentha piperita* essential oils. J. Agric. Food Chem. 2002, 50, 3943–3946.
- 83. Turgeon, T.; Wright, L. Mint. The Genus Mentha. J. Nat. Prod. 2007, 70, 1834.
- 84. Gochev, V.; Stoyanova, A.; Girova, T.; Atanasova, T. Chemical composition and antimicrobial activity of Bulgarian peppermint oils. *Bulg. Sci. Pap.* **2008**, *36*, 83–89.
- Zhang, L.; Xu, S.-G.; Liang, W.; Mei, J.; Di, Y.-Y.; Lan, H.-H.; Yang, Y.; Wang, W.-W.; Luo, Y.-Y.; Wang, H.-Z. Antibacterial Activity and Mode of Action of *Mentha arvensis* Ethanol Extract against Multidrug-Resistant Acinetobacter baumannii. *Trop. J. Pharm. Res.* 2015, 14, 2099.
- Bibi, S.; Ali, S.; Shahidin; Ullah, I.; Rauf, A.; Arif, D.; Umar, M.; Khan, A.S.; Hussain, M.; Ghani, A.; et al. Chemical composition and anti-microbial analysis of *Mentha arvensis* L. and *Thymus linearis* Benth. essential oils of leaves. *Rom. Biotechnol. Lett.* 2021, 26, 2893–2900. [CrossRef]

- Bokhari, N.; Perveen, K.; Al Khulaifi, M.; Kumar, A.; Siddiqui, I. In Vitro Antibacterial Activity and Chemical Composition of Essential Oil of *Mentha arvensis* Linn. Leaves. J. Essent. Oil-Bear. Plants 2016, 19, 907–915.
- Bassolé, I.H.N.; Lamien-Meda, A.; Bayala, B.; Tirogo, S.; Franz, C.; Novak, J.; Nebié, R.C.; Dicko, M.H. Composition and Antimicrobial Activities of *Lippia multiflora* Moldenke, *Mentha* × *piperita* L. and *Ocimum basilicum* L. Essential Oils and Their Major Monoterpene Alcohols Alone and in Combination. *Molecules* 2010, 15, 7825–7839.
- Tyagi, A.K.; Malik, A. Antimicrobial potential and chemical composition of *Mentha piperita* oil in liquid and vapour phase against food spoiling microorganisms. *Food Control* 2011, 22, 1707–1714. [CrossRef]
- Nikolić, M.; Jovanović, K.K.; Marković, T.; Marković, D.; Gligorijević, N.; Radulović, S.; Soković, M. Chemical composition, antimicrobial, and cytotoxic properties of five Lamiaceae essential oils. *Ind. Crops Prod.* 2014, 61, 225–232. [CrossRef]
- Iseppi, R.; Tardugno, R.; Brighenti, V.; Benvenuti, S.; Sabia, C.; Pellati, F.; Messi, P. Phytochemical Composition and In Vitro Antimicrobial Activity of Essential Oils from the Lamiaceae Family against *Streptococcus agalactiae* and *Candida albicans* Biofilms. *Antibiotics* 2020, 9, 592. [CrossRef]
- 92. Coutinho, H.D. Factors influencing the virulence of Candida spp. West Indian Med. J. 2009, 58, 160–163.
- 93. Benzaid, C.; Belmadani, A.; Djeribi, R.; Rouabhia, M. The Effects of *Mentha* × *piperita* Essential Oil on C. albicans Growth, Transition, Biofilm Formation, and the Expression of Secreted Aspartyl Proteinases Genes. *Antibiotics* 2019, *8*, 10.
- 94. Jayan, L.; Priyadharsini, N.; Ramya, R.; Rajkumar, K. Evaluation of antifungal activity of mint, pomegranate and coriander on fluconazole-resistant Candida glabrata. *J. Oral Maxillofac. Pathol.* **2020**, *24*, 517–522. [PubMed]
- Saharkhiz, M.J.; Motamedi, M.; Zomorodian, K.; Pakshir, K.; Hemyari, K.K. Chemical Composition, Antifungal and Antibiofilm Activities of the Essential Oil of *Mentha piperita* L. *ISRN Pharm.* 2012, 2012, 718645. [CrossRef] [PubMed]
- 96. Santos, K.K.; Matias, E.F.; Souza, C.E.; Tintino, S.R.; Braga, M.F.; Guedes, G.M.; Nogueira, L.F.; Morais, E.C.; Costa, J.G.; Menezes, I.R.; et al. Anti-Candida activity of *Mentha arvensis* and *Turnera ulmifolia*. J. Med. Food **2012**, 15, 322–324. [PubMed]
- Ibrahim, S.Y.; El-Salam, M.A. Anti-dermatophyte efficacy and environmental safety of some essential oils commercial and in vitro extracted pure and combined against four keratinophilic pathogenic fungi. *Environ. Health Prev. Med.* 2015, 20, 279–286. [PubMed]
- Tullio, V.; Roana, J.; Scalas, D.; Mandras, N. Evaluation of the Antifungal Activity of *Mentha* × *piperita* (Lamiaceae) of Pancalieri (Turin, Italy) Essential Oil and Its Synergistic Interaction with Azoles. *Molecules* 2019, 24, 3148. [PubMed]
- Scalas, D.; Roana, J.; Mandras, N.; Banche, G.; Allizond, V.; Cuffini, A.; Tullio, V. *Mentha* × *piperita* (Huds)(Lamiaceae) essential oil of Pancalieri (Turin, Italy): Preliminary evaluation of the antifungal activity and synergistic interaction with antifungal drugs. *Natural* 2015, 147, 61–62.
- 100. Nardoni, S.; Mugnaini, L.; Pistelli, L.; Leonardi, M.; Sanna, V.; Perrucci, S.; Pisseri, F.; Mancianti, F. Clinical and mycological evaluation of an herbal antifungal formulation in canine Malassezia dermatitis. *J. Mycol. Med.* **2014**, *24*, 234–240.
- Rachitha, P.; Krupashree, K.; Jayashree, G.V.; Gopalan, N.; Khanum, F. Growth Inhibition and Morphological Alteration of Fusarium sporotrichioides by *Mentha piperita* Essential Oil. *Pharmacogn. Res.* 2017, 9, 74–79.
- Camele, I.; Grul'ová, D.; Elshafie, H.S. Chemical Composition and Antimicrobial Properties of *Mentha* × *piperita* cv. 'Kristinka' Essential Oil. *Plants* 2021, 10, 1567.
- Diánez, F.; Santos, M.; Parra, C.; Navarro, M.J.; Blanco, R.; Gea, F.J. Screening of antifungal activity of 12 essential oils against eight pathogenic fungi of vegetables and mushroom. *Lett. Appl. Microbiol.* 2018, 67, 400–410.
- Scartazzini, L.; Tosati, J.V.; Cortez, D.H.C.; Rossi, M.J.; Flôres, S.H.; Hubinger, M.D.; Di Luccio, M.; Monteiro, A.R. Gelatin edible coatings with mint essential oil (*Mentha arvensis*): Film characterization and antifungal properties. *J. Food Sci. Technol.* 2019, 56, 4045–4056.
- 105. Pandey, A.K.; Rai, M.K.; Acharya, D. Chemical Composition and Antimycotic Activity of the Essential Oils of Corn Mint (*Mentha arvensis*) and Lemon Grass (*Cymbopogon flexuosus*) Against Human Pathogenic Fungi. *Pharm. Biol.* 2003, 41, 421–425. [CrossRef]
- Kaur, H.; Tandon, R.; Kalia, A.; Maini, C. Chemical composition and antifungal activity of essential oils from aerial parts of Mentha piperita and Mentha arvensis. Int. J. Pharmacol. 2018, 5, 767–773.
- 107. Stringhini, S.; Guessous, I. The Shift From Heart Disease to Cancer as the Leading Cause of Death in High-Income Countries: A Social Epidemiology Perspective. Ann. Intern. Med. 2018, 169, 877–878. [PubMed]
- 108. Pucci, C.; Martinelli, C.; Ciofani, G. Innovative approaches for cancer treatment: Current perspectives and new challenges. *Ecancermedicalscience* **2019**, *13*, 961. [CrossRef] [PubMed]
- Bhalla, Y.; Gupta, V.K.; Jaitak, V. Anticancer activity of essential oils: A review. J. Sci. Food Agric. 2013, 93, 3643–3653. [CrossRef]
 [PubMed]
- Nirmala, M.J.; Durai, L.; Anusha, G.S.; Nagarajan, R. Nanoemulsion of *Mentha arvensis* Essential Oil as an Anticancer Agent in Anaplastic Thyroid Cancer Cells and as an Antibacterial Agent in Staphylococcus aureus. *BioNanoScience* 2021, 11, 1017–1029. [CrossRef]
- Rahimifard, N.; Haji, M.H.; Hedayati, M.; Bagheri, O.; Pishehvar, H.; Ajani, Y. Cytotoxic effects of essential oils and extracts of some Mentha species on Vero, Hela and Hep2 cell lines. J. Med. Plants 2010, 9, 88–92.
- 112. Jain, D.; Pathak, N.; Khan, S.; Raghuram, G.V.; Bhargava, A.; Samarth, R.; Mishra, P.K. Evaluation of cytotoxicity and anticarcinogenic potential of Mentha leaf extracts. *Int. J. Toxicol.* 2011, 30, 225–236. [CrossRef] [PubMed]
- 113. Sonawane, H.; Shinde, A.; Jadhav, J. Evaluation of anti-angiogenic potential of *Mentha arvensis* Linn. Leaf extracts using chorioallantoic membrane assay. *World J. Pharm. Res.* **2016**, *5*, 677–689.

- 114. Kumar, A.; Samarth, R.M.; Yasmeen, S.; Sharma, A.; Sugahara, T.; Terado, T.; Kimura, H. Anticancer and radioprotective potentials of *Mentha piperita*. *Biofactors* **2004**, 22, 87–92.
- 115. Banerjee, P.P.; Bandyopadhyay, A.; Harsha, S.N.; Policegoudra, R.S.; Chattopadhyay, A. *Mentha arvensis* (Linn.)-mediated green silver nanoparticles trigger caspase 9-dependent cell death in MCF7 and MDA-MB-231 cells. *Breast Cancer* 2017, *9*, 265. [PubMed]
- 116. Weecharangsan, W.; Sithithaworn, W.; Siripong, P. Cytotoxic activity of essential oils of Mentha spp. on human carcinoma cells. *J. Health Res.* **2014**, *28*, 9–12.
- 117. Jerard, C.; Michael, B.P.; Chenicheri, S.; Vijayakumar, N.; Ramachandran, R. Rosmarinic Acid-Rich Fraction from Mentha arvensis Synchronizes Bcl/Bax Expression and Induces Go/G1 Arrest in Hepatocarcinoma Cells. *Proc. Natl. Acad. Sci. USA* 2019, 90, 515–522. [CrossRef]
- 118. Baskar, R.; Lee, K.A.; Yeo, R.; Yeoh, K.W. Cancer and radiation therapy: Current advances and future directions. *Int. J. Med. Sci.* **2012**, *9*, 193–199. [CrossRef] [PubMed]
- Baudino, T.A. Targeted Cancer Therapy: The Next Generation of Cancer Treatment. *Curr. Drug Discov. Technol.* 2015, 12, 3–20.
 [PubMed]
- 120. Citrin, D.E.; Mitchell, J.B. Altering the response to radiation: Sensitizers and protectors. Semin. Oncol. 2014, 41, 848–859. [CrossRef]
- 121. Cairnie, A.B. Adverse effects of radioprotector WR2721. Radiat. Res. 1983, 94, 221–226.
- Jagetia, G.C.; Baliga, M.S. Influence of the leaf extract of *Mentha arvensis* Linn. (mint) on the survival of mice exposed to different doses of gamma radiation. *Strahlenther. Onkol.* 2002, 178, 91–98.
- Samarth, R.M.; Goyal, P.K.; Kumar, A. Modulatory effect of *Mentha piperita* (Linn.) on serum phosphatases activity in Swiss albino mice against gamma irradiation. *Indian J. Exp. Biol.* 2001, 39, 479.
- 124. Samarth, R.M.; Goyal, P.K.; Kumar, A. Modulation of serum phosphatases activity in Swiss albino mice against gamma irradiation by *Mentha piperita* Linn. *Phytother. Res.* 2002, *16*, 586–589. [CrossRef] [PubMed]
- 125. Samarth, R.M.; Kumar, A. *Mentha piperita* (Linn.) leaf extract provides protection against radiation induced chromosomal damage in bone marrow of mice. *Indian J. Exp. Biol.* **2003**, *41*, 229–237. [PubMed]
- Bhatia, A.L.; Sharma, A.; Patni, S.; Sharma, A.L. Prophylactic effect of flaxseed oil against radiation-induced hepatotoxicity in mice. *Phytother. Res.* 2010, 21, 852–859. [CrossRef] [PubMed]
- 127. Highman, B.; Hanks, A.R. Serum intestinal alkaline phosphatase in rats after 800 R whole-body or regional x-irradiation. *Proc. Soc. Exp. Biol. Med.* **1970**, *133*, 1201–1206. [CrossRef]
- 128. Stepan, J.J.; Havránek, T.; Jojková, K. Serum alkaline phosphatases as indicators of radiation damage in rats. *Radiat. Res.* **1977**, *70*, 406–414.
- 129. Samarth, R.M.; Samarth, M. Protection against radiation-induced testicular damage in Swiss albino mice by *Mentha piperita* (Linn.). *Basic Clin. Pharmacol. Toxicol.* **2010**, 104, 329–334.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.